BMA BIOMEDICALS

Peninsula Laboratories

Buffer A
Extraction Wash buffer

Reconstitution Information:

This buffer is ready to use as a wash buffer in a solid phase extraction protocol using a C-18 Column (**# Y-1000**). Each 500 ml solution contains 1% trifluoroacetic acid in USP water. Store at 0 – 4 °C.

Recommended Use: **Extraction of Peptide from Plasma**

1. Add an equal amount of Buffer A to the plasma. Example: 1 ml of plasma with 1 ml of Buffer A. Centrifuge at 6,000xg to 17,000xg for 20 min at 4°C. Discard pellet.
2. Equilibrate a Sep-Column (**# Y-1000**) with Buffer B **(#Y-1045**) (or 100% Acetonitrile) (1x 1.0 ml,) followed by Buffer A (3x 3.0 ml).
3. Load the plasma solution onto the pre-treated Sep-Column.

   Note: For steps 4 and 5, a light vacuum (10 sec per drop) may be applied to the column.
4. Slowly wash the column with Buffer A (2x 3.0 ml), discard the wash.
5. Elute the peptide slowly with Buffer B (1x 3.0 ml), collect eluant in a PP tube.
6. Evaporate eluant to dryness using a centrifugal concentrator or by a suitable method.
7. Dissolve the residue in RIA buffer **(#Y-1050**) for radioimmunoassay: For a normal subject, dissolve in 250 µl buffer for a two-tube assay. Aliquot 100 µl into each tube. If each tube is found to contain 3.963 pg/tube, then the total level of peptide in the sample = 3.962 pg/tube x 2.5 tube = 9.9 pg. If upon assaying, the peptide value exceeds or does not fall in the range of detection, dilute or concentrate the samples accordingly.
8. The total time for extraction should be 0.5 days and 0.5 days for lyophilization. After lyophilization, store the sample at -70°C before assaying, analyze as soon as possible.

NOTE: This is a generic extraction protocol, which can be used for multiple types of samples. If a more suitable extraction method is found in literature, we encourage the researcher to use it.

This product contains no preservative and is intended for laboratory use and research purposes only. Not for diagnostic or therapeutic applications.

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